

Protein L Magnetic Beads

CATALOG #: 6537-1

AMOUNT: 1 ml

LOT #: _____

PREPARATION: Protein L Magnetic Beads are prepared by covalently coupling Recombinant Protein L (contains five Ig kappa light chain binding domains, BV catalog # 6530) to 6% cross-linked magnetically beaded agarose. The coupling technique is optimized to give a high binding capacity. The capacity of IgG binding is generally greater than 10 mg of human IgG per ml of wet gel.

CONTENTS: Supplied as a 50% slurry in 20 % Ethanol/dW.

TECHNICAL SPECIFICATIONS:

Parameter	Description
Support Characteristics	Paramagnetic, spherical, 6 % cross-linked agarose
Ligand	Recombinant Protein L
Particle Size	75 – 150 µm
Binding Capacity	Generally >10 mg human IgG/ml wet beads
Working Temperature	Room temperature
Storage Solution	20 % Ethanol/dW
Storage Temperature	4 – 8 °C
Stability	Stable, as supplied, for at least 1 year

FEATURES: Easy to use, high-binding capacity, non-adherent beads. Useful for immunoprecipitation and enrichment of antibodies. High affinity for kappa-light chain containing Ig antibodies from a variety of species. Protein L binds to all IgG subclasses from human, mouse and rat species. It also binds to human, mouse and rat IgM, IgA, IgE and IgD, as well as FAB with Kappa-light chains. Protein L is superior for binding chicken, hamster and pig IgG.

FOR RESEARCH USE ONLY! Not to be used on humans.

SUGGESTED PROTOCOL:

Prepare the antibody solution by diluting the required amount of antibody in binding buffer before running the protocol.

1. Magnetic Bead Preparation (perform three times)
 - a. Dispense the required amount of magnetic beads into a 1.5 ml microfuge tube.
 - b. Place the tube in the magnetic rack and remove the storage solution.
 - c. Add 500 µl binding buffer.
 - d. Resuspend the beads.
 - e. Remove the liquid
2. Antibody Capture
 - a. Immediately add the antibody solution.
 - b. Resuspend and mix (slow end-over-end) for at least 15 minutes.
 - c. Remove the liquid.
3. Washing
 - a. Add 500 µl Binding Buffer containing 0.5 M NaCl; Remove the liquid.
 - b. Add 500 µl Binding Buffer; Remove the liquid.
4. Target Binding
 - a. Add sample diluted in binding buffer.
 - b. Incubate with slow end-over-end mixing for up to 60 minutes.
 - c. Remove and collect unbound fraction.
5. Washing (perform three times)
 - a. Add 500 µl wash buffer
 - b. Remove liquid (save washes to troubleshoot)
6. Elution (perform three times)
 - a. Add 2 volumes elution buffer (vs. bead volume).
 - b. Completely resuspend beads and incubate at least 2 minutes.
 - c. Remove and collect elution fraction.

RECOMMENDED BUFFER EXAMPLES:

- Binding buffer:** 50 mM Tris, 150 mM NaCl, pH 7.5
- Wash buffer:** 50 mM Tris, 150 mM NaCl, pH 7.5 (or add 1% Octylglucoside to this buffer)
(Could also try 1X PBS as both binding and wash buffer)
- Elution buffer:** 0.1 M -0.2 M Glycine pH 2.5-3.1 (or 0.1 M citric acid, pH 2.5-3.1 or 2.5 % Acetic Acid)

RELATED PRODUCTS:

Recombinant Protein A	Protein A Sepharose	Protein A Magnetic Beads
Recombinant Protein G	Protein G Sepharose	Protein G Magnetic Beads
Recombinant Protein L	Protein L Sepharose	Protein L Magnetic Beads
Recombinant Protein A/G	Protein A/G Sepharose	Protein A/G Magnetic Beads
Recombinant Protein A/G/L	Protein A/G/L Sepharose	Protein A/G/L Magnetic Beads
Protein G Polyclonal Antibody	Protein G-Biotin	Protein G-FITC
Protein A Polyclonal Antibody	Protein G Coated Plates	
Protein L Polyclonal Antibody		