



EnzyChrom[™] Fructose Assay Kit (EFRU-100)

Quantitative Colorimetric Fructose Determination at 565nm

DESCRIPTION

FRUCTOSE (C₆H₁₂O₆, also called levulose or laevulose), is a monosaccharide found in honey, tree fruits, berries, melons, and some root vegetables along with glucose and galactose. The human body can use fructose for energy, however, too much consumption may lead to high triglycerides. Simple, direct and high-throughput assays for fructose determination find wide applications. BioAssay Systems' reagent systems reacts directly and specifically with fructose to form a colored product. Glucose and galactose do not interfere. The color intensity at 565nm is directly proportional to the fructose concentration in the sample.

KEY FEATURES

Use as little as 20 μL samples. Linear detection range in 96-well plate: 12 to 1000 μM fructose.

APPLICATIONS

Direct Assays: fructose in biological samples (e.g. serum, plasma, urine, saliva, milk, culture medium), food, juice, beverage and other agricultural products.

Drug Discovery/Pharmacology: effects of drugs on fructose metabolism.

KIT CONTENTS

Standard: 400 µL 20 mM D-Fructose

Storage conditions. The kit is shipped on ice. Store all components at -20°C. Shelf life of three months after receipt.

Precautions: reagents are for research use only. Normal precautions for laboratory reagents should be exercised while using the reagents. Please refer to Material Safety Data Sheet for detailed information.

ASSAY PROCEDURE

Note: (1) The following substances interfere and should be avoided in sample preparation: ascorbic acid, SDS (>0.2%), sodium azide, NP-40 (>1%) and Tween-20 (>1%). (2) This assay is based on a kinetic reaction. To ensure identical incubation time, addition of Working Reagent to standard and samples should be quick and mixing should be brief but thorough. Use of a multi-channel pipettor is recommended.

Sample treatment: liquid samples such as serum, plasma and fruit juices can be assayed directly. Because fruit juices may contain high concentrations of fructose, it is recommended to dilute juice sample 50-fold (n=50) in dH₂O prior to assay. Milk samples should be cleared by mixing 600 μ L milk with 100 μ L 6 N HCl. Centrifuge 5 min at 14,000 rpm. Transfer 300 μ L supernatant into a clean tube and neutralize with 50 μ L 6 N NaOH. The neutralized supernatant is ready for assay (dilution factor n=1.36).

- Equilibrate all components to room temperature. Briefly centrifuge the tubes before opening. Keep thawed tubes on ice during assay.
- 2. Standards: mix 12 μL 20 mM Standard with 228 μL dH₂O (final 1000 μM). Dilute standard in dH₂O as follows.

No	1000 μM STD + H ₂ O	Vol (μL)	Fructose (μM)
1	100 μL + 0 μL	100	1000
2	60 μL + 40 μL	100	600
3	30 μL + 70 μL	100	300
4	0 μL +100 μL	100	0

Transfer 20 μL diluted standards into separate wells of a clear flat-bottom 96-well plate.

Samples: transfer 20 μL of each sample into separate wells of the plate.

3. Color reaction. Prepare enough Working Reagent by mixing, for each reaction well, 56 μ L Assay Buffer, 1 μ L Enzyme, 14 μ L PMS Solution and 14 μ L MTT Solution.

Keep Working Reagent protected from light. Add 80 μ L Working Reagent to each well. Tap plate to mix. *Do not expose Working Reagent to light for more than 5 minutes.* Incubate 60 min at room temperature in the dark.

4. Read optical density at 565nm (520-600nm).

Note: If the calculated fructose concentration of a sample is higher than 1000 μ M, dilute sample in water and repeat the assay. Multiply result by the dilution factor *n*.

CALCULATION

Subtract blank value (water, #4) from the standard values and plot the ΔOD against standard concentrations. Determine the slope and calculate the fructose concentration of Sample,

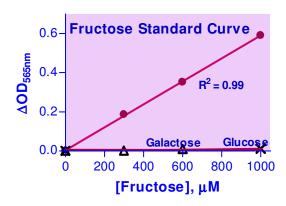
[Fructose] =
$$\frac{OD_{Sample} - OD_{H2O}}{Slope (\mu M^{-1})} \times n \quad (\mu M)$$

 $\mathsf{OD}_{\mathsf{SAMPLE}},\,\mathsf{OD}_{\mathsf{H2O}}$ are optical density values of the sample and water. n is the dilution factor.

Conversions: 1 mM fructose equals 18 mg/dL, 0.018% or 180 ppm.

MATERIALS REQUIRED, BUT NOT PROVIDED

Pipetting devices, centrifuge tubes, clear flat-bottom uncoated 96-well plates, optical density plate reader.



LITERATURE

- 1. Novelli G, Reichardt JK. (2000). Molecular basis of disorders of human fructose metabolism: past, present, and future. Mol Genet Metab. 71:62-65.
- 2. Pudek MR et al. (1990). Low concentration fructose determination in plasma adapted to the Cobas-Bio. Clin Biochem. 23:221-223.
- 3. Gabrielli M. (1978). Serum fructose determination with centrifugal analyzers. Clin. Chem. 24:1990-1995.

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