

Human Factor V ELISA Kit

Introduction

Factor V (FV) is an essential cofactor of the blood coagulation cascade and circulates in plasma as a large single-chain glycoprotein (330 kDa). The deduced amino acid sequence consists of 2224 amino acids inclusive of a 28-amino acid leader peptide (1). During coagulation, it is converted to the active cofactor FVa via limited proteolysis by thrombin, and spliced into a heavy chain (110 KDa) and a light chain (73 KDa) held together non-covalently by calcium (2). In the presence of a calcium ion and the phospholipid on cell surfaces, FVa and FXa form the prothrombinase complex which catalyzes the hydrolysis of prothrombin to thrombin (3). Thrombin in turn cleaves fibrinogen to fibrin which polymerizes to form a clot. FVa is readily inactivated by anticoagulant activated protein C (4). FV Leiden, a single amino acid mutation, renders FVa resistant to cleavage by activated protein C. It therefore over-produces thrombin, and leads to excess clotting and hereditary thrombophilia disorder (5). Severe FV deficiency is associated with mild to severe bleeding diathesis (6).

Principal of the Assay

The Human Factor V ELISA kit is designed for detection of human Factor V in plasma, urine, milk, and cell culture supernatants. This assay employs a quantitative sandwich enzyme immunoassay technique that measures Factor V in less than 4 hours. A polyclonal antibody specific for Factor V has been pre-coated onto a microplate. Factor V in standards and samples is sandwiched by the immobilized antibody and a biotinylated polyclonal antibody specific for Factor V, which is recognized by a streptavidin-peroxidase conjugate. All unbound material is then washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

Caution and Warning

- Prepare all reagents (working diluent buffer, wash buffer, standards, biotinylated-antibody, and SP conjugate) as instructed, prior to running the assay.
- Prepare all samples prior to running the assay. The dilution factors for the samples are suggested in this protocol. However, the user should determine the optimal dilution factor.
- Spin down the SP conjugate vial and the biotinylated-antibody vial before opening and using contents.
- This kit is for research use only.
- The kit should not be used beyond the expiration date.
- The Stop Solution is an acid solution.

Reagents

- **Factor V Microplate:** A 96-well polystyrene microplate (12 strips of 8 wells) coated with a monoclonal antibody against Factor V.
- **Sealing Tapes:** Each kit contains 3 pre-cut, pressure-sensitive sealing tapes that can be cut to fit the format of the individual assay.
- Factor V Standard: Human Factor V in a buffered protein base (60 ng, lyophilized).
- **Biotinylated Factor V Antibody (80x):** A 80-fold concentrated biotinylated polyclonal antibody against human Factor V (100 µl).
- EIA Diluent Concentrate (10x): A 10-fold concentrated buffered protein base (30 ml).
- Wash Buffer Concentrate (20x): A 20-fold concentrated buffered surfactant (30 ml, 2 bottles).
- Streptavidin-Peroxidase Conjugate (SP Conjugate): A 100-fold concentrate (80 µl).
- **Chromogen Substrate**: A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- Stop Solution: A 0.5 N hydrochloric acid to stop the chromogen substrate reaction (12 ml).

Storage Condition

- Store components of the kit at 2-8°C or -20°C upon arrival up to the expiration date.
- Store SP Conjugate and Biotinylated Antibody at -20^oC
- Store Microplate, Diluent Concentrate (10x), Wash Buffer, Stop Solution, and Chromogen Substrate at 2-8°C
- Opened unused microplate wells may be returned to the foil pouch with the desiccant packs. Reseal along zip-seal. May be stored for up to 1 month in a vacuum desiccator.
- Diluent (1x) may be stored for up to 1 month at 2-8°C. Store Standard at 2-8°C before reconstituting with Diluent and at -20°C after reconstituting with Diluent

Other Supplies Required

- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes (1-20 µl, 20-200 µl, 200-1000 µl and multiple channel)
- Deionized or distilled reagent grade water

Sample Collection, Preparation and Storage

- **Plasma:** Collect plasma using one-tenth volume of 0.1 M sodium citrate as an anticoagulant. Centrifuge samples at 2000 x g for 10 minutes. Dilute samples 1:800 with EIA Diluent. Store the remaining samples at -20°C or below. Avoid repeated freeze-thaw cycles. (EDTA or Heparin can also be used as anticoagulant.)
- **Cell Culture Supernatants:** Centrifuge cell culture media at 2000 x g for 10 minutes to remove debris. Collect supernatants and assay. Store the remaining samples at -20°C or below. Avoid repeated freeze-thaw cycles.
- **Urine:** Collect urine using sample pot. Centrifuge samples at 600 x g for 10 minutes. Urine dilution is suggested at 1:2 in EIA Diluent. Store samples at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.
- Milk: Collect milk using sample tube. Centrifuge samples at 600 x g for 10 minutes. Milk dilution is suggested at 1:2 in EIA Diluent. Store samples at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.

Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use. If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.
- **EIA Diluent Concentrate (10x):** Dilute EIA Diluent Concentrate 1:10 with reagent grade water. Store for up to 1 month at 2-8°C.
- **Standard Curve:** Reconstitute the 60 ng of Human Factor V Standard with 1 ml of EIA Diluent to generate a stock solution of 60 ng/ml. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare duplicate or triplicate standard points by serially diluting the Factor V standard solution (60 ng/ml) 1:2 with EIA Diluent to produce 30, 15, 7.5, 3.75, 1.875, and 0.938 ng/ml solutions. EIA Diluent serves as the zero standard (0 ng/ml). Any remaining solution should be frozen at -20°C.

Standard Point	Dilution	[FV] (ng/ml)
P1	1 part Standard (60 ng/ml)	60.000
P2	1 part P1 + 1 part EIA Diluent	30.000
P3	1 part P2 + 1 part EIA Diluent	15.000
P4	1 part P3 + 1 part EIA Diluent	7.500
P5	1 part P4 + 1 part EIA Diluent	3.750
P6	1 part P5 + 1 part EIA Diluent	1.875
P7	1 part P6 + 1 part EIA Diluent	0.938
P8	EIA Diluent	0.000

- **Biotinylated Factor V Antibody (80x):** Spin down the antibody briefly and dilute the desired amount of the antibody 1:80 with EIA Diluent. Any remaining solution should be frozen at -20°C.
- Wash Buffer Concentrate (20x): Dilute Wash Buffer Concentrate 1:20 with reagent grade water
- **SP Conjugate (100x):** Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with EIA Diluent. Any remaining solution should be frozen at -20°C.

Assay Procedure

- Prepare all reagents, working standards and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-30°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccant inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.
- Add $50~\mu l$ of Standard or sample per well. Cover wells and incubate for two hours. Start the timer after the last sample addition.
- Wash five times with 200 µl of Wash Buffer manually. Invert the plate each time and decant the contents; hit it 4-5 times on absorbent paper towel to completely remove the liquid. If using a machine wash six times with 300 µl of Wash Buffer and then invert the plate, decant the contents; hit it 4-5 times on absorbent paper towel to completely remove the liquid.
- Add 50 µl of Biotinylated Factor V Antibody to each well and incubate for one hour.
- Wash the microplate as described above.
- Add 50 μl of Streptavidin-Peroxidase Conjugate per well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.

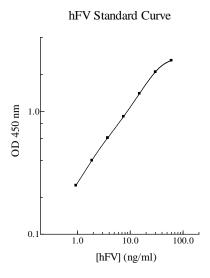
- Wash the microplate as described above.
- Add 50 µl of Chromogen Substrate per well and incubate for about 20 minutes or till the optimal blue color density develops. Gently tap the plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 µl of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm **immediately**. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections. Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

Data Analysis

- Calculate the mean value of the duplicate or triplicate readings for each standard and sample.
- To generate a standard curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit.
- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

Standard Curve

• The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.



Performance Characteristics

- The minimum detectable level of Factor V is typically ~ 0.9 ng/ml.
- Intra-assay and inter-assay coefficients of variation were 4.8 % and 7.4% respectively.

Linearity

	Average Percentage of Expected Value	
Sample Dilution	Plasma	
1:400	95%	
1:800	98%	
1:1600	93%	

	Average Percentage of Expected Value	
Sample Dilution	Urine	Milk
No dilution	96%	92%
1:2	97%	95%
1:4	91%	89%

Recovery

Standard Added Value	2-20 ng/ml
Recovery %	82-111 %
Average Recovery %	96%

Cross-Reactivity

Species	% Cross Reactivity	
Canine	1%	
Bovine	None	
Monkey	20% (suggested 1:40 dilution for plasma)	
Mouse	0.1%	
Rat	0.1%	
Swine	40% (suggested 1:100 dilution for plasma)	
Rabbit	None	

References

- (1) Jenny RJ et al. (1987) Proc. Natl. Acad. Sci. U.S.A. 84:4846-4850
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- (4) Kisiel W et al. (1977) Biochemistry 16:5824-5831
- (5) Bertina RM et al. (1994) Nature 369:64-67
- (6) Asselta R et al. (2006) J. Thromb. Haemost 4(1):26-34

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