Human Protein C ELISA Kit (Plasma and Serum Samples)

Introduction

Protein C is a vitamin K-dependent plasma antithrombotic and anti-inflammatory zymogenic glycoprotein that is synthesized in the liver. Protein C has a light chain of 155 amino acids (21 kDa) and a heavy chain of 262 amino acids (41 kDa) linked by a disulfide bond. On endothelial cell membrane, thrombin-thrombomodulin complex cleaves a 12-reside peptide from protein C amino terminus of the heavy chain and converts it to activated protein C (APC). APC inactivates coagulation Factor Va and Factor VIIIa and performs a major role in regulating blood clotting, inflammation, and apoptosis (1-3). Protein C deficiency causes neonatal purpura fulminans, thrombophilia, and recurrent venous thrombosis (4-6). Protein C pathway components have been studied in the treatment of complex disorders, including severe sepsis, thrombosis, and ischemic stroke (7).

Principal of the Assay

The Human Protein C ELISA (Enzyme-Linked Immunosorbent Assay) kit is designed for detection of human Protein C in plasma and serum. This assay employs a quantitative competitive enzyme immunoassay technique that measures human Protein C in less than 3 hours. A polyclonal antibody specific for human Protein C has been pre-coated onto a 96-well microplate with removable strips. Protein C in standards and samples is competed with a biotinylated Protein C sandwiched by the immobilized antibody and streptavidin-peroxidase conjugate. All unbound material is then washed away and a peroxidase enzyme substrate is added. The color development is stopped and the intensity of the color is measured.

Caution and Warning

- Prepare all reagents (working diluent buffer, wash buffer, standards, biotinylated-protein, and SP conjugate) as instructed, prior to running the assay.
- Prepare all samples prior to running the assay. The dilution factors for the samples are suggested in this protocol. However, the user should determine the optimal dilution factor.
- Spin down the SP conjugate vial before opening and using contents.
- This kit is for research use only.
- The kit should not be used beyond the expiration date.
- The Stop Solution is an acid solution

Reagents

- **Human Protein C Microplate:** A 96-well polystyrene microplate (12 strips of 8 wells) coated with a polyclonal antibody against human Protein C.
- **Sealing Tapes:** Each kit contains 3 pre-cut, pressure-sensitive sealing tapes that can be cut to fit the format of the individual assay.
- **Human Protein** C **Standard:** Human Protein C in a buffered protein base (6 μg, lyophilized).
- **Biotinylated Protein C:** 1 vial, lyophilized.
- EIA Diluent Concentrate (10x): A 10-fold concentrated buffered protein base (30 ml).
- Wash Buffer Concentrate (20x): A 20-fold concentrated buffered surfactant (30 ml).
- Streptavidin-Peroxidase Conjugate (SP Conjugate): A 100-fold concentrate (80 µl).
- **Chromogen Substrate**: A ready-to-use stabilized peroxidase chromogen substrate tetramethylbenzidine (8 ml).
- Stop Solution: A 0.5 N hydrochloric acid to stop the chromogen substrate reaction (12 ml).

Storage Condition

- Store components of the kit at 2-8°C or -20°C upon arrival up to the expiration date.
- Store SP Conjugate at -20^oC
- Store Microplate, Diluent Concentrate (10x), Wash Buffer, Stop Solution, and Chromogen Substrate at 2-8°C
- Opened unused microplate wells may be returned to the foil pouch with the desiccant packs. Reseal along zip-seal. May be stored for up to 1 month in a vacuum desiccator.
- Diluent (1x) may be stored for up to 1 month at $2-8^{\circ}$ C.
- Store Standard and Biotinylated Protein at 2-8°C before reconstituting with Diluent and at -20°C after reconstituting with Diluent.

Other Supplies Required

- Microplate reader capable of measuring absorbance at 450 nm.
- Pipettes (1-20 μl, 20-200 μl, 200-1000μl and multiple channel).
- Deionized or distilled reagent grade water.

Sample Collection, Preparation and Storage

- **Plasma:** Collect plasma using one-tenth volume of 0.1 M sodium citrate as an anticoagulant. Centrifuge samples at 2000 x g for 10 minutes. Dilute samples 1:8 into EIA Diluent. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles (EDTA or Heparin can also be used as anticoagulant).
- **Serum:** Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 2000 x g for 10 minutes. Remove serum and assay. Dilute samples 1:8 into EIA Diluent. The undiluted samples can be stored at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles.

Reagent Preparation

- Freshly dilute all reagents and bring all reagents to room temperature before use. If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.
- **EIA Diluent Concentrate (10x):** Dilute the EIA Diluent 1:10 with reagent grade water. Store for up to 1 month at 2-8°C.

• Standard Curve: Reconstitute the 6 μg of Protein C Standard with 1 ml of EIA Diluent to generate a solution of 6 μg/ml. Allow the standard to sit for 10 minutes with gentle agitation prior to making dilutions. Prepare duplicate or triplicate standard points by serially diluting the standard solution (6 μg/ml) 1:2 with equal volume EIA Diluent to produce 3, 1.5, 0.75, 0.375, 0.188 and 0.094 μg/ml solutions. EIA Diluent serves as the zero standard (0 μg/ml). Any remaining solution should be frozen at -20°C.

Standard Point	Dilution	[Protein C] (µg/ml)
P1	Standard (6 µg/ml)	6.000
P2	1 part P1 + 1 part EIA Diluent	3.000
P3	1 part P2 + 1 part EIA Diluent	1.500
P4	1 part P3 + 1 part EIA Diluent	0.750
P5	1 part P4 + 1 part EIA Diluent	0.375
P6	1 part P5 + 1 part EIA Diluent	0.188
P7	1 part P6 + 1 part EIA Diluent	0.094
P8	EIA Diluent	0.000

- **Biotinylated Protein C** (1x): Dilute Biotinylated Protein C with 4 ml EIA Diluent to produce a working solution. Allow to sit for 10 minutes with gentle agitation prior to use. Any remaining solution should be frozen at -20°C.
- Wash Buffer Concentrate (20x): Dilute the Wash Buffer Concentrate 1:20 with reagent grade water.
- **SP Conjugate** (100x): Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 1:100 with EIA Diluent. Any remaining solution should be frozen at -20°C.

Assay Procedure

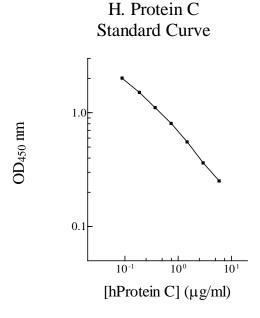
- Prepare all reagents, working standards and samples as instructed. Bring all reagents to room temperature before use. The assay is performed at room temperature (20-30°C).
- Remove excess microplate strips from the plate frame and return them immediately to the foil pouch with desiccant inside. Reseal the pouch securely to minimize exposure to water vapor and store in a vacuum desiccator.
- Add 25 µl of standard or sample per well, and immediately add 25 µl of Biotinlylated Protein C to each well (on top of the Standard or sample) and mix gently. Cover wells with a sealing tape and incubate for two hours. Start the timer after the last sample addition.
- Wash five times with 200 µl of Wash Buffer manually. Invert the plate each time and decant
 the contents; hit it 4-5 times on absorbent paper towel to completely remove the liquid.
 If using a machine wash six times with 300 µl of Wash Buffer and then invert the plate,
 decant the contents; hit it 4-5 times on absorbent paper towel to completely remove the
 liquid.
- Add 50 µl of Streptavidin-Peroxidase Conjugate to each well and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
- Wash the microplate as described above.
- Add 50 µl of Chromogen Substrate per well and incubate for about 10 minutes or till the optimal blue color density develops. Gently tap plate to ensure thorough mixing and break the bubbles in the well with pipette tip.
- Add 50 µl of Stop Solution to each well. The color will change from blue to yellow.
- Read the absorbance on a microplate reader at a wavelength of 450 nm **immediately**. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections. Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

Data Analysis

- Calculate the mean value of the duplicate or triplicate readings for each standard and sample.
- To generate a Standard Curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance on the y-axis. The best-fit line can be determined by regression analysis using log-log or four-parameter logistic curve-fit.
- Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

Standard Curve

• The curve is provided for illustration only. A standard curve should be generated each time the assay is performed.



Performance Characteristics

- The minimum detectable dose of Protein C is typically 0.9 µg/ml.
- Intra-assay and inter-assay coefficients of variation were 4.8% and 7.3% respectively.

Linearity

	Average Percentage of Expected Value	
Sample Dilution	Plasma	Serum
1:4	95%	105%
1:8	97%	100%
1:16	89%	91%

Recovery

Standard Added Value	0.2-2 ug/ml
Recovery %	81-108 %
Average Recovery %	96 %

Cross-Reactivity

Species	% Cross Reactivity
Canine	10%
Bovine	20%
Monkey	50%
Mouse	2%
Rat	20%
Swine	100%
Human	100%
Rabbit	80%

Reference Value

• The normal blood level of Protein C1 is $\sim 4 \mu g/ml$.

References

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