SensiFAST™ SYBR & Fluorescein One-Step Kit

Shipping: On Dry/Blue Ice

Exp. Date: See vial

Batch No.: See vial

Concentration: see vial

Catalog Numbers

BIO-75001: 100 x 20µl reactions: 1 x 1ml

BIO-75005: 500 x 20µl reactions: 5 x 1ml

BIOLINE

A Meridian Life Science® Company

Store at –20°C

Storage and Stability:
The SensiFAST SYBR & Fluorescein One-Step Kit is shipped on Dry/Blue Ice. All kit components should be stored at -20°C upon receipt. Excessive freeze/thawing is not recommended. When stored under optimum conditions, the reagents are stable for a minimum of 12 months from date of purchase.

Quality Control:
Bioline operates under ISO 9001 Management System. The SensiFAST SYBR & Fluorescein One-Step Kit and its components are extensively tested for activity, processivity, efficiency, heat activation, sensitivity and absence of nuclease contamination and nucleic acid contamination.

Safety Precautions:
Harmful if swallowed. Irritating to eyes, respiratory system and skin. Please refer to the material safety data sheet for further information.

Notes:
For research use only.

Description

The SensiFAST™ SYBR & Fluorescein One-Step Kit has been formulated for highly reproducible first-strand cDNA synthesis and subsequent real-time PCR in a single tube. A combination of the latest advances in buffer chemistry together with a reverse transcriptase and hot-start DNA polymerase system ensures that SensiFAST SYBR & Fluorescein One-Step Kit produces fast, highly-specific and ultra-sensitive one-step RT-qPCR.

The SensiFAST SYBR & Fluorescein One-Step Kit consists of a 2x SensiFAST SYBR One-Step mix, as well as separate reverse transcriptase and RiboSafe RNase Inhibitor.

Kit components

Reagent	100 x 20µl Reactions	500 x 20µl Reactions
SensiFAST™ SYBR & Fluorescein One-Step mix (2x)	1 x 1ml	5 x 1ml
RiboSafe RNase Inhibitor	1 x 40µl	1 x 200µl
Reverse transcriptase	1 x 20µl	1 x 100µl
DEPC-H_2O	1 x 1.8ml	2 x 1.8ml

Instrument compatibility

SensiFAST SYBR & Fluorescein One-Step Kit has been optimized for use in SYBR Green-based real-time RT-PCR on the real-time instruments listed in the following compatibility table, each of these instruments having the capacity to analyze the real-time PCR data with the passive reference signal either on or off. The kit is also compatible with several instruments that do not require the use of fluorescein, such as the Qiagen (Corbett) Rotor-Gene™ 6000, the Bio-Rad CFX96 or the Roche LightCycler® 480.

Manufacturer	Model
Bio-Rad	iCycler™, MyiQ™, iQ™5

General considerations

When handling RNA, it is important to use RNase-free plasticware and reagents. We also recommend performing RNA work in an isolated area. To help prevent any carry-over DNA contamination we recommend that separate areas are maintained for reaction set-up, PCR amplification and any post-PCR gel analysis. It is essential that any tubes containing amplified PCR product are not opened in the PCR set-up area.

Primers: The sequence and concentration of the primers, as well as amplicon length, can be critical for specific amplification, yield and overall efficiency of any RT-qPCR. We strongly recommend taking the following points into consideration when designing and running your RT-qPCR:

- use primer-design software, such as Primer3 or visual OMP™ (http://frodo.wi.mit.edu/primer3/ and DNA Software, Inc. http://dnasoftware.com/, respectively). Primers should have a melting temperature (Tm) of approximately 60°C
- optimal amplicon length should be 80-200bp, and should not exceed 400bp
- final primer concentration of 400nM is suitable for most SYBR-Green based reactions, however to determine the optimal concentration we recommend titrating in the range 0.1-1µM
- use an equimolar primer concentration
- where possible, use intron-spanning primers to avoid amplification from genomic DNA

Template: It is important that the RNA template is intact and devoid of DNA or contaminating inhibitors of both reverse transcription and PCR. For high purity RNA, we recommend using the Bioline ISOLATE RNA Mini Kit (BIO-52043). RNA stocks and dilutions should be made in DEPC-treated Water (BIO-38030) to avoid any RNase-mediated degradation.

The recommended amount of template for one-step RT-qPCR is dependent upon the type of RNA used.

- **total RNA:** purified total RNA can be used in the range from 1pg to 1µg per 20µl reaction
- **mRNA:** purified mRNA can be used from 0.01pg per 20µl reaction

$MgCl_2$: The $MgCl_2$ concentration in the 1x reaction mix is 3mM. In the majority of RT-qPCR conditions this is optimal for both the reverse transcriptase and the hot-start DNA polymerase. If necessary, we suggest titrating the $MgCl_2$ to a maximum of 5mM.

RT-PCR controls: It is important to detect the presence of contaminating DNA that may affect the reliability of the data. Always include a no-RT control by omitting the reverse transcriptase from the reaction.

Optional Fluorescein well-factor correction: SYBR Fluorescein Kit is premixed with fluorescein, so that fluorescence emitted by fluorescein can be optionally detected on certain real-time instruments. If your real-time instrument has the capability of using fluorescein and you wish to use this option, then this option must be selected by the user in the software (*see notice to purchaser No. 5 in Trademark and Licensing Information*).

Procedure

Reaction mix composition: Prepare a RT-PCR mastermix. The volumes given below are based on a standard 20µl final reaction mix and can be scaled accordingly.

Reagent	Volume	Final concentration
2x SensiFAST™ SYBR & Fluorescein One-Step Mix	10µl	1x
10µM Forward Primer	0.8µl	400nM
10µM Reverse Primer	0.8µl	400nM
Reverse transcriptase	0.2µl	-
RiboSafe RNase Inhibitor	0.4µl	-
H_2O	up to 16µl	
Template	4µl	
	20µl Final volume	

Suggested RT-qPCR conditions: The following RT-qPCR conditions are suitable for the SensiFAST SYBR & Fluorescein One-Step Kit with the majority of amplicons and real-time PCR instruments. However, the cycling conditions can be varied to suit different machine-specific protocols. SensiFAST SYBR & Fluorescein One-Step Kit is compatible with either 3-step or 2-step cycling:

- **3-step cycling**

Cycles	Temperature	Time	Notes
1	45°C	10min	Reverse transcription
1	95°C	2min	Polymerase activation
40	95°C 60°C 72°C	5s 10s 5s	Denaturation Annealing Extension (acquire at end of step)

- **2-step cycling**

Cycles	Temperature	Time	Notes
1	45°C	10min	Reverse transcription
1	95°C	2min	Polymerase activation
40	95°C 60°C	5s 20s	Denaturation Annealing/extension (acquire at end of step)

Optional analysis: After the reaction has reached completion, refer to the instrument instructions for the option of melt-profile analysis.

Troubleshooting Guide

Problem	Possible Cause	Recommendation
No amplification trace AND No product on agarose gel	Activation time too short	Ensure SensiFAST SYBR One-Step mix is activated for a minimum of 2min at 95°C before cycling
	Error in protocol setup	Verify that correct reagent concentrations, volumes, dilutions and storage conditions have been used
	Suboptimal primer design	Use primer design software or validated assay. Test assay on a control template
	Incorrect concentration of primers	Use primer concentrations between 100nM and 1µM
	Template degraded	Re-isolate your template from the sample material or use freshly prepared template dilution. We recommend using the ISOLATE RNA kits for template preparation and DEPC-treated water for resuspension or dilution of the template Verify the integrity of RNA using agarose gel electrophoresis
	Primers degraded	Use newly synthesized primers
	Template contaminated with RT-PCR inhibitors	Further dilute template before RT-PCR or purify template and resuspend it in DEPC-treated water
	Template concentration too low	Increase concentration used
	Cycling conditions not optimal	Increase extension/annealing time, increase cycle number, reduce annealing temperature

Troubleshooting Guide (Continued)

Problem	Possible Cause	Recommendation
No amplification trace AND PCR product present on agarose gel	Error in instrument setup	Check that the acquisition settings are correct during cycling
Non-specific amplification product AND Primer-dimers	Inefficient reverse transcription	Extend reverse transcription time up to 20min and/or increase the temperature up to 48°C
	Suboptimal primer design	Redesign primers using appropriate software, or use validated primers
	Primer concentration too high	Test dilution series of primer concentrations until primer-dimer/non-specific amplification products disappear
	Primer concentration too low	Use primer concentration between 100nM and 1µM
	Primer annealing temperature too low	Increase PCR annealing temperature up to 65°C or until primer-dimer/non-specific amplification products disappear
	Template concentration too low	Increase template concentration
	Template concentration too high	Reduce template concentration until non-specific products disappear
	Extension time too long	Reduce extension time to determine whether non-specific products are reduced
Variability between replicates	Error in reaction set-up	Prepare large volume mastermix
	Air bubbles in reaction mix	Centrifuge reaction samples/plate prior to running on a real-time instrument
Late amplification trace	Inefficient reverse transcription	Extend reverse transcription time up to 20min and/or increase the temperature up to 48°C
	Activation time too short	Ensure SensiFAST SYBR & Fluoresein One-Step mix is activated for a minimum of 1min at 95°C before cycling
	Annealing temperature too high	Decrease annealing temperature in steps of 2°C
	Extension time too short	Double extension time to determine whether the cycle threshold (C_T) is affected
	Template concentration too low	Increase concentration if possible
	Template is degraded	Re-isolate template from sample material or use freshly prepared template dilution
	Suboptimal primer design	Redesign primers using appropriate software, or use validated primers
	Primer concentration too low	Increase concentration of primers in 100nM increments
	RNase contamination	Ensure RNase inhibitor is added before addition of template
PCR efficiency below 90%	Extension time too short	Increase extension time
	Primer concentration too low	Increase concentration of primers in 100nM increments
	Suboptimal primer design	Redesign primers using appropriate software or use validated primers
PCR efficiency above 110%	Template is degraded or contains PCR inhibitors	Re-isolate template from sample material, or use freshly prepared template dilution, or purify template and resuspend it in water
	Non-specific amplification and/or primer -dimers	Use 4% agarose gel electrophoresis to confirm presence of non-specific amplification products. See above for preventing/removing non-specific products

Associated Products

Product	Description	Pack Size	Cat No.
ISOLATE Genomic DNA Mini kit	Rapid isolation of DNA from a variety of samples	10 Preps 50 Preps 250 Preps	BIO-52031 BIO-52032 BIO-52033
ISOLATE Plant DNA Mini kit	Rapid isolation of DNA from a variety of plant samples	10 Preps 50 Preps 250 Preps	BIO-52034 BIO-52035 BIO-52036
ISOLATE RNA Mini Kit	Fast and efficient isolation of extremely pure total RNA from a variety of samples	10 Preps 50 Preps 250 Preps	BIO-52039 BIO-52040 BIO-52041
ISOLATE Plant RNA Mini Kit	Fast and efficient isolation of extremely pure total RNA from a variety of plant samples	10 Preps 50 Preps 250 Preps	BIO-52042 BIO-52043 BIO-52044
TRIsure™	Quick isolation of high-quality RNA from a variety of sources for subsequent use in cDNA synthesis	100ml 200ml	BIO-38032 BIO-38033
cDNA Synthesis Kit	Fully optimized to generate maximum yields of full-length cDNA from RNA	30 Reactions 100 Reactions	BIO-65025 BIO-65026
Agarose	Molecular biology grade agarose	100g 500g	BIO-41026 BIO-41025
PCR Water	Ultra-pure (18.2MΩ) molecular biology grade water	10 x 10ml	BIO-37080
DEPC-treated Water	Deionized, high-quality molecular grade water treated with DEPC. Ideal for use in all RNA work	10 x 10ml 1 Liter	BIO-38030 BIO-38031

Technical Support

If the troubleshooting guide does not solve the difficulty you are experiencing, please contact Technical Support with details of reaction setup, cycling conditions and relevant data.

Email: tech@bioline.com

Trademark and Licensing Information

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