

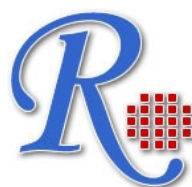
RayBio[®]
Human/Mouse/Rat Inhibin B
(Beta B subunit)
Enzyme Immunoassay Kit

**Please Read the Manual Carefully
Before Starting your Experiment**

**User Manual 2.2
(Revised March 15, 2012)**

**RayBio[®] Inhibin B Enzyme
Immunoassay Kit Protocol**

(Cat#: EIA-INB-1)



RayBiotech, Inc.

**We Provide You With Excellent
Protein Array System and Service**

**Tel: (Toll Free) 1-888-494-8555 or 770-729-2992; Fax: 770-206-2393;
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**RayBio® Human/Mouse/Rat Inhibin B (beta B subunit)
Enzyme Immunoassay Kit Protocol**

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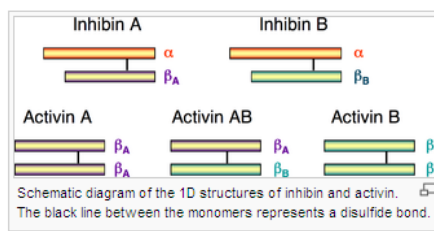
I. INTRODUCTION

Inhibin B belongs to TGF- β superfamily, which members also include Inhibin A, TGF-betas (transforming growth factors), activins, BMPs (bone morphogenetic proteins). Among these factors, inhibins are most closely related to activins.

The activin and inhibin are both dimeric protein complexes in structure, and in each complex, the two monomers are linked to one another by a single disulfide bond. In addition, both complexes are derived from the same family of related genes and proteins but differ in their subunit composition. Inhibins are disulfide-linked heterodimers comprising an alpha-subunit and beta-subunit.

Inhibin B is composed of one alpha subunit and one beta B subunit; Inhibin A is composed of one alpha subunit and one beta A subunit. By contrast, activins are homo- or hetero-dimers, forming activin A (2 beta A subunits), activin B (2 beta B subunits), or activin AB (1 beta A subunit and 1 beta B subunit).

Class	Activity	Complex	Dimer subunits	
			1	2
Inhibin	inhibits FSH secretion	Inhibin A	α	β_A
		Inhibin B	α	β_B
Activin	stimulates FSH secretion	Activin A	β_A	β_A
		Activin AB	β_A	β_B
		Activin B	β_B	β_B



Inhibins have played important roles in regulating reproductive health. Inhibins and activins generally play opposing roles as modulators of the reproductive axis and have multiple roles in the regulation of reproductive physiology in both females and males. Recently the emerging roles for inhibins in tumorigenesis have been reported. Inhibins has been reported as potential tumor suppressors in several different tissues, including ovary, prostate, endometrium and breasts, with loss of inhibin expression or loss of sensitivity linked to tumor initiation and progression as well as poor patient survival.

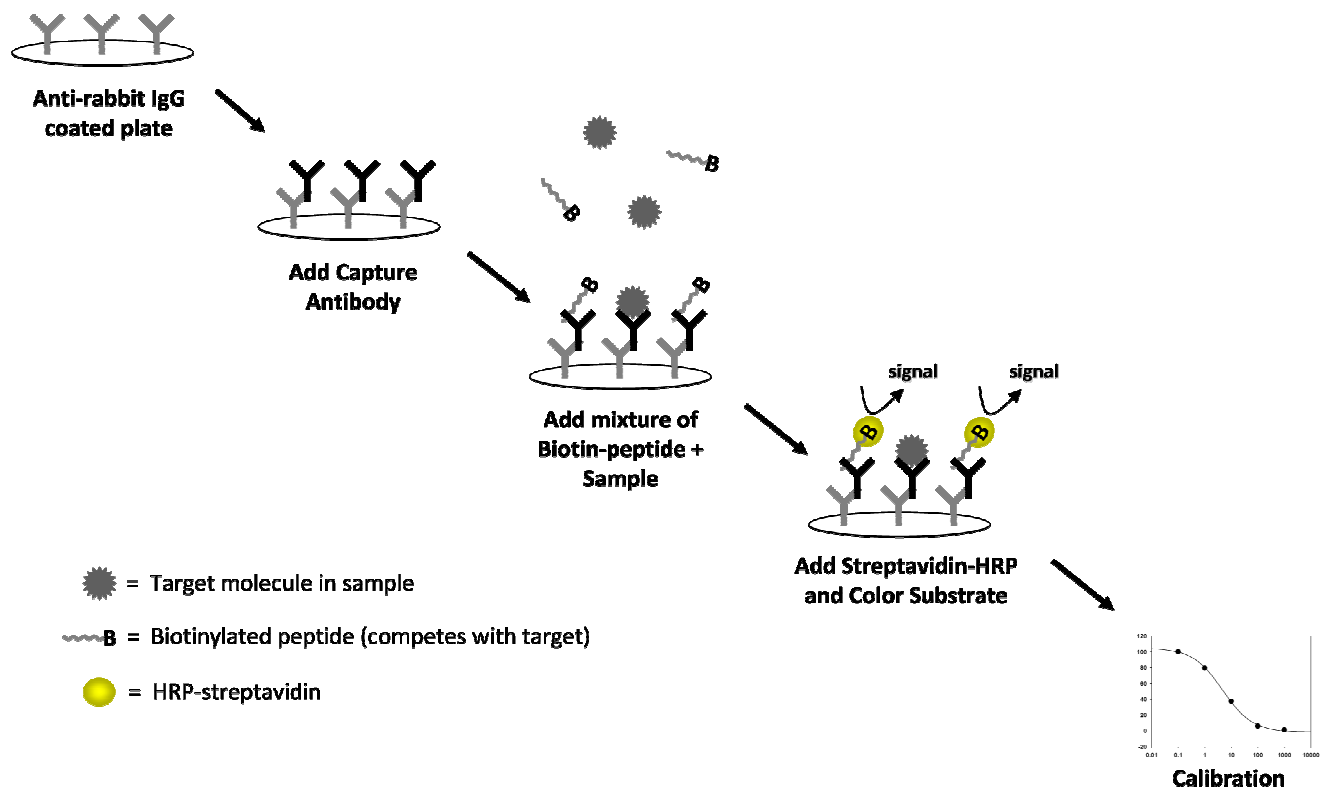
II. GENERAL DESCRIPTION

The RayBio® Inhibin B Enzyme Immunoassay (EIA) Kit is an in vitro quantitative assay for detecting Inhibin B peptide based on the principle of Competitive Enzyme Immunoassay.

The microplate in the kit is pre-coated with anti-rabbit secondary antibody. After a blocking step and incubation of the plate with anti-Inhibin B antibody, both biotinylated Inhibin B peptide and peptide standard or targeted peptide in samples interacts competitively with the Inhibin B antibody. Uncompeted (bound) biotinylated Inhibin B peptide then interacts with Streptavidin-horseradish peroxidase (SA-HRP), which catalyzes a color development reaction. The intensity of colorimetric signal is directly proportional to the amount of biotinylated peptide-SA-HRP complex and inversely proportional to the amount of Inhibin B peptide in the standard or samples. This is due to the competitive binding to Inhibin B antibody between biotinylated Inhibin B peptide and peptides in standard or samples. A standard curve of known concentration of Inhibin B peptide can be established and the concentration of Inhibin B peptide in the samples can be calculated accordingly.

EIA-INB-1 targets the beta B subunit, and therefore theoretically detects Activin B and Activin AB in addition to Inhibin B.

Principle of Competitive EIA



III. REAGENTS

1. Inhibin B Microplate (Item A): 96 wells (12 strips x 8 wells) coated with secondary antibody.
2. Wash Buffer Concentrate (20x) (Item B): 25 ml
3. Standard Inhibin B Peptide (Item C): 2 vials, 10 µl/vial
4. Anti-Inhibin B polyclonal antibody (Item N): 2 vials, 5 µl/vial
5. Assay Diluent A (Item D): 30 ml, contains 0.09% sodium azide as preservative. Diluent for standards and serum or plasma samples.
6. Assay Diluent B (Item E): 15 ml of 5x concentrated buffer. Diluent for standards and cell culture media or other sample types.
7. Biotinylated Inhibin B peptide, (Item F): 2 vials, 20 µl/vial
8. HRP-Streptavidin concentrate (Item G): 600 µl 400x concentrated HRP-conjugated Streptavidin.
9. Positive control (Item M): 1 vial, 100 µl
10. TMB One-Step Substrate Reagent (Item H): 12 ml of 3, 3', 5, 5'- tetramethylbenzidine (TMB) in buffered solution.
11. Stop Solution (Item I): 8 ml of 0.2 M sulfuric acid.
12. Assay Diagram (Item J).
13. User Manual (Item K)

IV. STORAGE

- Standard, Biotinylated Inhibin B peptide, and Positive Control should be stored at -20 °C or -80 °C (recommended at -80 °C) after arrival. **Avoid multiple freeze-thaws.**
- The remaining kit components may be stored at -20 °C.
- Opened Microplate Wells and antibody (Item N) may be stored for up to 1 month at 2° to 8 °C. Return unused wells to the pouch containing desiccant pack and reseal along entire edge.
- If stored in this manner, RayBiotech warrants this kit for 6 months from the date of shipment.

V. ADDITIONAL MATERIALS REQUIRED

1. Microplate reader capable of measuring absorbance at 450nm.
2. Precision pipettes to deliver 2 µl to 1 ml volumes.
3. Adjustable 1-25 ml pipettes for reagent preparation.
4. 100 ml and 1 liter graduated cylinders.
5. Absorbent paper.
6. Distilled or deionized water.
7. SigmaPlot software (or other software which can perform four-parameter logistic regression models)
8. Tubes to prepare standard or sample dilutions.
9. Orbital shaker
10. Aluminum foil
11. Saran Wrap

VI. REAGENT PREPARATION

If testing plasma or serum samples, use Assay Diluent A to dilute Item F and Item C. If testing cell culture media or other sample types, use Assay Diluent B to dilute Item F and Item C. For sample and positive control dilutions, refer to steps 6, 7, 8 and 10 of Reagent Preparation.

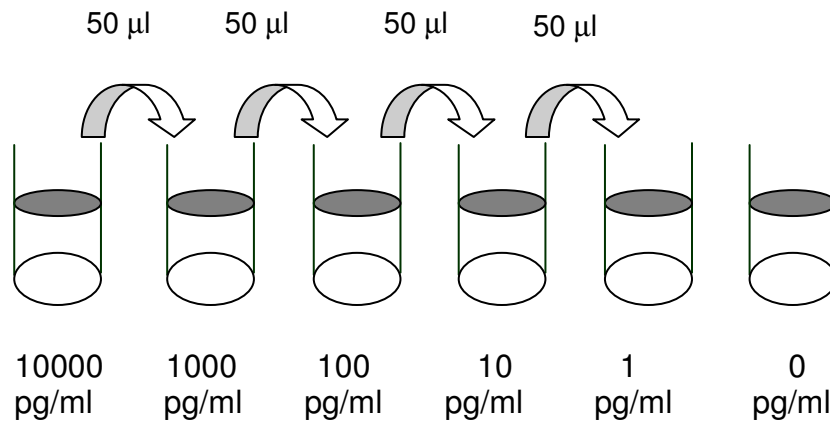
1. Keep kit reagents on ice during reagent preparation steps. Equilibrate plate to room temperature before opening the sealed pouch.
2. Assay Diluent B (Item E) should be diluted 5-fold with deionized or distilled water.
3. Briefly centrifuge the Anti-Inhibin B Antibody vial (Item N) before use. Add 50 µl of 1x Assay Diluent B into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently.

4. The antibody concentrate should then be diluted 100-fold with 1x Assay Diluent B. This is your anti-Inhibin B antibody working solution, which will be used in step 2 of the Assay Procedure.

NOTE: the following steps may be done during the antibody incubation procedure (step 2 of Assay Procedure).

5. Briefly centrifuge the vial of Biotinylated Inhibin B (Item F) before use. Add 5 µl of Item F to 5 ml of the appropriate Assay Diluent. Pipette up and down to mix gently. *The final concentration of biotinylated Inhibin B will be 100 pg/ml.* This solution will only be used as the diluent in step 6 of Reagent Preparation.
6. Preparation of Standards: Label 6 microtubes with the following concentrations: 10000 pg/ml, 1000 pg/ml, 100 pg/ml, 10 pg/ml, 1 pg/ml and 0 pg/ml. Pipette 450 µl of biotinylated Inhibin B solution into each tube, except for the 10000 pg/ml (leave this one empty). *It is very important to make sure the concentration of biotinylated Inhibin B is 100 pg/ml in all standards.*
 - a. Briefly centrifuge the vial of Inhibin B (Item C). In the tube labeled 10000 pg/ml, pipette 8 µl of Item C and 792 µl of 100 pg/ml biotinylated Inhibin B solution (prepared in step 5 above). This is your Inhibin B stock solution (10000 pg/ml Inhibin B, 100 pg/ml biotinylated Inhibin B). Mix thoroughly. This solution serves as the first standard.
 - b. To make the 1000 pg/ml standard, pipette 50 µl of Inhibin B stock solution the tube labeled 1000 pg/ml. Mix thoroughly.
 - c. Repeat this step with each successive concentration, preparing a dilution series as shown in the illustration below. Each time, use 450 µl of biotinylated Inhibin B and 50 µl of the prior concentration until 1 pg/ml is reached. Mix each tube thoroughly before the next transfer.

d. The final tube (0 pg/ml Inhibin B, 100 pg/ml biotinylated Inhibin B) serves as the zero standard (or total binding).



7. Prepare a 10-fold dilution of Item F. To do this, add 2 µl of Item F to 18 µl of the appropriate Assay Diluent. This solution will be used in steps 8 and 10.
8. Positive Control Preparation: briefly centrifuge the positive control vial (Item M). To the tube of Item M, add 101 µl 1x Assay Diluent B. Also add 2 µl of 10-fold diluted Item F (prepared in step 7) to the tube. This is a 2-fold dilution of the positive control. Mix thoroughly. The positive control is a cell culture medium sample with an expected signal between 10% and 30% of total binding (70-90% competition) if diluted as described above. It may be diluted further if desired, but be sure the final concentration of biotinylated Inhibin B is 100 pg/ml.
9. If Item B (20X Wash Concentrate) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1X Wash Buffer.

10. Sample Preparation: Use Assay Diluent A + biotinylated Inhibin B to dilute serum/plasma samples. For cell culture medium and other sample types, use 1X Assay Diluent B + biotinylated Inhibin B as the diluent. *It is very important to make sure the final concentration of the biotinylated Inhibin B is 100 pg/ml in every sample.* EXAMPLE: to make a 4-fold dilution of sample, mix together 2.5 µl of 10-fold diluted Item F (prepared in step 7), 185 µl of appropriate Assay Diluent, and 62.5 µl of your sample; mix gently. The total volume is 250 µl, enough for duplicate wells on the microplate.
Do not use Item F diluent from Step 5 for sample preparation. If you plan to use undiluted samples, you must still add biotinylated Inhibin B to a final concentration of 100 pg/ml. EXAMPLE: Add 2.5 µl of 10-fold diluted Item F to 247.5 µl of sample. NOTE: Optimal sample dilution factors should be determined empirically, however you may contact technical support (888-494-8555; techsupport@raybiotech.com) to obtain recommended dilution ranges for serum or plasma.
11. Briefly centrifuge the HRP-Streptavidin vial (Item G) before use. The HRP-Streptavidin concentrate should be diluted 400-fold with 1X Assay Diluent B.

Note: Do not use Assay Diluent A for HRP-Streptavidin preparation in Step 11.

VII. ASSAY PROCEDURE:

1. Keep kit reagents on ice during reagent preparation steps. It is recommended that all standards and samples be run at least in duplicate.
2. Add 100 µl anti-Inhibin B antibody (see Reagent Preparation step 4) to each well. Incubate for 1.5 hours at room

temperature with gentle shaking (1-2 cycles/sec). You may also incubate overnight at 4 degrees C.

3. Discard the solution and wash wells 4 times with 1x Wash Buffer (200-300 μ l each), Washing may be done with a multichannel pipette or an automated plate washer. Complete removal of liquid at each step is essential to good assay performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
4. Add 100 μ l of each standard (see Reagent Preparation step 6), positive control (see Reagent Preparation step 8) and sample (see Reagent Preparation step 10) into appropriate wells. Be sure to include a blank well (Assay Diluent only). Cover wells and incubate for 2.5 hours at room temperature with gentle shaking (1-2 cycles/sec) or overnight at 4°C.
5. Discard the solution and wash 4 times as directed in Step 3.
6. Add 100 μ l of prepared HRP-Streptavidin solution (see Reagent Preparation step 11) to each well. Incubate for 45 minutes with gentle shaking at room temperature. It is recommended that incubation time should not be shorter or longer than 45 minutes.
7. Discard the solution and wash 4 times as directed in Step 3.
8. Add 100 μ l of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with gentle shaking (1-2 cycles/sec).
9. Add 50 μ l of Stop Solution (Item I) to each well. Read absorbances at 450 nm immediately.

VIII. ASSAY PROCEDURE SUMMARY

1. Prepare all reagents, samples and standards as instructed.



2. Add 100 μ l anti-Inhibin B antibody to each well. Incubate 1.5 hours at room temperature or overnight at 4°C.



3. Add 100 μ l standard or sample to each well. Incubate 2.5 hours at room temperature or overnight at 4°C.



4. Add 100 μ l prepared streptavidin solution. Incubate 45 minutes at room temperature.



5. Add 100 μ l TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.



6. Add 50 μ l Stop Solution to each well. Read at 450 nm immediately.

IX. CALCULATION OF RESULTS

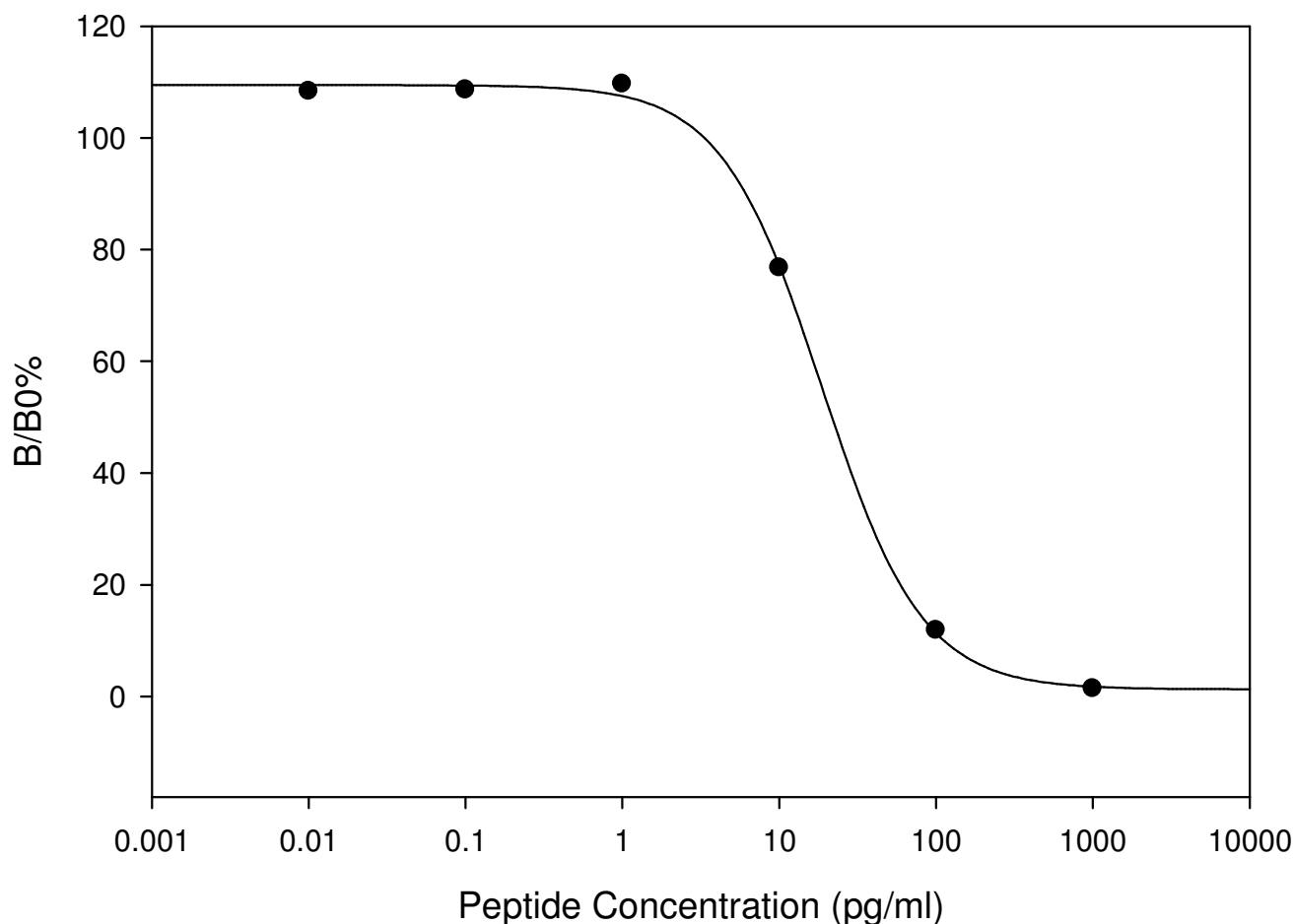
Calculate the mean absorbance for each set of duplicate standards, controls and samples, and subtract the blank optical density. Plot the standard curve using SigmaPlot software (or other software which can perform four-parameter logistic regression models), with standard concentration on the x-axis and percentage of absorbance (see calculation below) on the y-axis. Draw the best-fit curve through the standard points.

Percentage absorbance = $(B - \text{blank OD}) / (B_0 - \text{blank OD})$ where
B = OD of sample or standard and
B₀ = OD of zero standard (total binding)

A. TYPICAL DATA

These standard curves are for demonstration only. A standard curve must be run with each assay.

2D Graph 1



B. SENSITIVITY

The minimum detectable concentration of Inhibin B is 34.6 pg/ml.

C. DETECTION RANGE

1-10,000 pg/ml

D. REPRODUCIBILITY

Intra-Assay: CV<10%

Inter-Assay: CV<15%

X. SPECIFICITY

Cross Reactivity: This ELISA kit shows no cross-reactivity with any of the cytokines tested: Ghrelin, Nesfatin, Angiotensin II, NPY and APC.

XI. REFERENCES

1. Burger HG, Igarashi M (1988). "Inhibin: definition and nomenclature, including related substances". *The Journal of Clinical Endocrinology and Metabolism* 66 (4): 885–6.
2. Robertson DM, Burger HG, Fuller PJ (2004). "Inhibin/activin and ovarian cancer". *Endocrine-related Cancer* 11 (1): 35–49.
3. Ying SY (December 1987). "Inhibins and activins: chemical properties and biological activity". *Proc. Soc. Exp. Biol. Med.* 186 (3): 253–64.

XII. TROUBLESHOOTING GUIDE

Problem	Cause	Solution
1. Poor standard curve	1. Inaccurate pipetting 2. Improper standard dilution	1. Check pipettes 2. Ensure briefly spin the vial of Item C and dissolve the powder thoroughly by a gentle mix.
2. Low signal	1. Too brief incubation times 2. Inadequate reagent volumes or improper dilution	1. Ensure sufficient incubation time; assay procedure step 2 change to over night 2. Check pipettes and ensure correct preparation
3. Large CV	1. Inaccurate pipetting	1. Check pipettes
4. High background	1. Plate is insufficiently washed 2. Contaminated wash buffer	1. Review the manual for proper wash. If using a plate washer, check that all ports are unobstructed. 2. Make fresh wash buffer
5. Low sensitivity	1. Improper storage of the EIA kit 2. Stop solution	1. Store your standard at $\leq -20^{\circ}\text{C}$ after receipt of the kit. 2. Stop solution should be added to each well before measure

RayBio® EIA kits:

If you are interested in other EIA kits, please visit www.raybiotech.com for details.

Notes:

This product is for research use only.



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3607 Parkway Lane, Suite 200
Norcross, GA 30092
Tel: 770-729-2992, 1-888-494-8555
Fax: 770-206-2393
Web: www.raybiotech.com