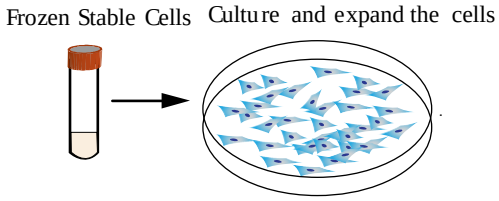
DATA SHEET

P53 Overexpression Stable Cell Line

Catalog Number SL-0100 **(For Research Use Only)**

Introduction

p53 plays a key role in cellular homeostasis and is at the heart of a complex network of protective mechanisms safeguarding cellular integrity. Because of its central function in processes such as cell cycle regulation, DNA repair, cellular senescence, and apoptosis, the p53 pathway is crucial for effective tumor suppression. Signosis has established a stable cell line with constitutive overexpression of wild type p53 with myc tag in Cos-7 cells. Therefore, the cell line can be used as an expression system for monitoring the function of of p53 in different applications, such as activation and protein-protein interactions.



Stable cell line diagram

Principle of the assay

The cell line was established by transfection of CMV p53 myc tag expression vector along with hygromycin expression vector followed by hygromycin selection. The hygromycin resistant clones were subsequently screened for p53 and myc tag expression. The clone with the highest expression was selected and expanded to produce this stable cell line.

Materials provided

- One vial of 5 X 10^6 cells, at passage 3, in Freezing Media (store the vial in liquid nitrogen until it is ready to be thawed).

Material required but not provided

- Dulbecco's Modified Eagle's Medium (DMEM)
- Fetal Bovine Serum (FBS)
- Penicillin (10,000 units/ml)
- Hygromycin B (Roche)
- Freezing media
- Luciferase reporter system (Promega E-1500)

Handling cells upon arrival

- It is strongly recommended that you propagate the cells by following instructions as soon as possible upon arrival.
- Genetic instability is a common in all transfected cells, therefore, it is critical to prepare numbers of frozen stocks at early passages.
- Prepare **Initial Growth Media**:
  DMEM (in high glucose + sodium pyruvate + L-glutamine + Phenol Red) + Penicillin (100 units/mL) Streptomycin (100ug/ml) + 10% FBS)
- Prepare **Complete Growth Media:**
  DMEM (in high glucose + sodium pyruvate + L-glutamine + Phenol Red) + Penicillin (100 units/mL) Streptomycin (100ug/ml) + 10% FBS + Hygromycin (100ug/ml)

Initial Culture Procedure

Important: The first propagation of cells should be for generating stocks for future use. Cells undergo genotypic changes resulting in reduced responsiveness over time in normal cell culture conditions. Therefore, it is critical to prepare an adequate number of frozen stocks at early passages.

1. Prepare culture dish by adding 15ml of pre-warmed **Initial Growth Media** to a 100-mm culture dish.
2. Quickly thaw cells in a 37 ^{o}C water bath with constant agitation.
3. Immediately transfer entire contents of the vial to the prepared culture dish. DO NOT pipette cells up and down as this may damage the cells.
4. Rock the culture dish to equally distribute the cells.
5. Place the culture dish with cells in a humidified incubator at 37 ^{o}C or 5% CO2.
6. After 48 hours, change to **Complete Growth Media**.
7. Change media every 2-3 days using Complete Growth Media.
8. When cells reach 90% confluency (usually within 1 week), prepare frozen stocks and continue propagate the rest of the culture for future assays.
9. Transfer vials to liquid nitrogen for long term storage.

Prepare frozen stocks

1. Carefully remove the culture media from cells by aspiration.
2. Rinse cells with PBS, being careful to not dislodge attached cells. Then remove PBS by aspiration.
3. Add 2ml of 0.25% Trypsin/0.53mM Tris-EDTA solution to the culture dish.
4. Let the dish incubate with Trypsin for a few minutes (2-3 min). Confirm detachment by observation under the microscope.
5. Add 10ml of pre-warmed Complete Growth Media and gently pipette up and down to break the clumps.
6. Transfer cells to a 15ml conical centrifuge tube and centrifuge at 125 x g for 5 minutes to collect the cells.
7. Aspirate the culture media and resuspend cells at a density of 5 x 10^6 cells/mL in freezing media.
8. Aliquot 1ml cells into cryogenic vials.
9. Place vials in a freezing container and store at -80^{o}C overnight.

Assay procedure for co-immunoprecipitation (IP)

The following procedure should be followed as a guideline. You will need to optimize the assay conditions based upon your experimental set up.

1. The day before performing the assay, trypsinize the cells and plate on a 10cm^2 dish with 70% confluency and incubate for overnight.
2. Wash the cells with 1XPBS once, add 400ul of cell lysis buffer and incubate for 1 hour one ice with gently shaking.
3. Collect the cells and centrifuge cell lysate at 12,000 rpm for 10 minutes.
4. The supernatant are ready for co-IP assay.
   The kit CI-0003 p53 co-IP kit can be used for the application.

Data Example

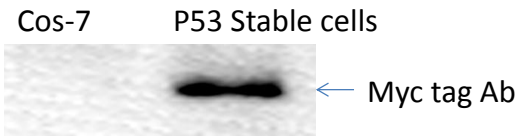


Figure2. Western blot analysis p53 Cos-7 Stable Cell line with myc tag antibody.